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Chemical Name: Afidopyropen

USEPA PC Code: 026200 USEPA MRID: 49689225 USEPA DP Barcode: 435146 PMRA Data Code (DACO): 9.2.4.8 PMRA Study No. (UKID): 2627490 Data Requirement: Non-guideline

Test Material: BAS 440 00 I (TEP, VERSYS™) **Purity: 9.7%**

Active Ingredient: Afidopyropen

IUPAC Name: [(3*S*,4*R*,4a*R*,6*S*,6a*S*,12*R*,12a*S*,12b*S*)-3-(cyclopropylcarbonyloxy)-1,2,3,4,4a,5,6,6a,12a,12b-decahydro-6,12-dihydroxy-4,6a,12b-trimethyl-11-oxo-9-(3pyridyl)-11H,12H-benzo[f]pyrano[4,3-b]chromen-4-yl]methylcyclopropane carboxylate

CAS Name: [(3*S*,4*R*,4a*R*,6*S*,6a*S*,12*R*,12a*S*,12b*S*)-3-(cyclopropylcarbonyl)oxy)]-

1,3,4,4a,5,6,6a,12,12a,12b-decahydro-6,12-dihydroxy-4,6a,12b-trimethyl-11-oxo-9-(3-

pyridyl)-2*H*,11*H*-naphtho[2,1-*b*]pyrano[3,4-*e*]pyran-4-yl]methyl

cyclopropanecarboxylate CAS No.: 915972-17-7 Synonyms: INSCALIS™

Primary Reviewer: Thomas Steeger, Ph.D.

Senior Science Advisor, USEPA/OSCP/OPP/EFED/ERB 4

Secondary Reviewer: Cameron Douglass, Ph.D.

Biologist, USEPA/OSCP/OPP/EFED/ERB 4

PMRA Reviewer: Vedad Izadi

Evaluation Officer, PMRA/EAD/ERSII

Date Evaluation Completed: 1 September 2017

CITATION: Klinkert, A. 2015. Determination of residues of BAS 440 00 I (Afidopyropen) in bee-

> relevant matrices in a semi-field honey bee (Apis mellifera L.) study in Canola (Brassica sp.) after one application during flowering. Eurofins Agroscience Services Eco Chem GmbH Eutinger Str. 24 75223 Niefern-Öschelbronn, Germany. Report No. 721406.

Sponsored by BASF.

Executive Summary:

A semi-field test was carried out to determine the residues of a typical end-use product (TEP) containing afidopyropen (BAS 440 00I, VERSYS™), and its metabolite M440I007, in flowers, pollen and nectar collected by honey bees (Apis mellifera) from canola (Brassica sp.) either treated once with BAS 440 00 I (9.7% active ingredient) or with water (negative control) during bloom at BBCH 63-64 in North Carolina, USA. BAS 440 00 I was applied as one application nominally corresponding to 50 g a.i./ha (0.272 lbs

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a.i./A) during active bee flight during flowering. All applications were performed by using a backpack boom-sprayer at a rate of 500 L water/ha.

Commercial honey bees were used as sampling device for canola pollen and nectar. Honeybee hives were set up in the day before application and first sampling. Samples of honey bee-collected pollen (pollen traps), nectar (honey stomach) collected from forager bees and canola flowers were taken 4 times and pollen directly from flowers once until end of flowering, were analyzed for residues of BAS 440 I (afidopyropen) and its metabolite M440I007.

In canola flowers, mean afidopyropen residues were 4.43 mg/kg (range: 3.73 - 4.97 mg/kg) at 0 days after application (0 DAA), 0.11 mg/kg (range: 0.10 - 0.13 mg/kg) at 3 DAA, and 0.013 mg/kg (range: 0.011 - 0.015 mg/kg) at 7 DAA, and were less than the limit of quantification (LOQ<0.01 mg/kg) at 10 DAA. Mean M440I007 residues were 0.36 mg/kg (range: 0.33 - 0.40 mg/kg) at 0 DAA, 0.020 mg/kg (range: 0.014 - 0.023 mg/kg) at 3 DAA, and were below the LOQ (<0.01 mg/kg) in all samples collected 7 and 10 DAA.

In pollen, mean afidopyropen residues were 0.26 mg/kg (range: 0.17 - 0.40 mg/kg) at 0 DAA, were 0.023 mg/kg (range, 0.018 - 0.031 mg/kg) and 0.027 mg/kg (range: 0.020 - 0.037 mg/kg) at 3 and 7 DAA, respectively, and declined to <0.010 mg/kg (range: <0.010 - 0.011 mg/kg) at 10 DAA. Mean M4401007 residues were 0.061 mg/kg (range: 0.037 - 0.10) at 0 DAA and were below the LOQ (each replicate <0.01 mg/kg, except one at 0.018 mg/kg at 3 DAA) for the remaining sampling intervals.

In nectar, mean afidopyropen residues were 0.052 mg/kg (range, 0.010 - 0.13 mg/kg) at 0 DAA, and were below the LOQ (<0.01 mg/kg) in all samples collected 3, 7, and 10 DAA. Mean M440I007 residues were below the LOQ (each replicate <0.01 mg/kg except one at 0.013 mg/kg at 0 DAA) in all samples collected 0, 3, 7, and 10 DAA.

Although the study was not intended to provide biological information on the bees used to collect samples, weights of pollen samples provide anecdotal information that at 0 and 3 DAA, samples sizes from treated plots were 77 – 78% lower than controls; however, by 7 and 10 DAA, pollen sample weights from treated plots had increased by 38 and 36%, respectively, relative to controls. These data suggest that bee pollen foraging activity may have been affected in afidopyropen-treated plots from 0 through 3 DAA; however, after this period, bee pollen foraging activity may have increased in the treated group relative to controls. Based on sample sizes of nectar, nectar foraging activity did not appear to be affected.

Results Synopsis:

Mean residues at 0 DAA after foliar application of BAS 440 00 I at nominal rate of 50 g a.i./ha (0.272 lbs a.i./A):

Flowers: 4.43 ± 0.63 mg/kg (Parent); 0.36 ± 0.035 mg/kg (M4401007) Nectar: 0.052 ± 0.068 mg/kg (Parent); <0.01 mg/kg (M4401007) Pollen: 0.26 ± 0.12 mg/kg (Parent); 0.061 ± 0.034 mg/kg (M4401007)

EPA Classification: Acceptable

PMRA Classification: Reliable with restrictions

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I. DATA SOURCE

USEPA MRID No.: 49689225 **PMRA UKID:** 2627490

Study Title: Determination of residues of BAS 440 00 I (Afidopyropen) in bee-

relevant matrices in a semi-field honey bee (Apis mellifera L.) study in

Canola (*Brassica* sp.) after one application during flowering.

Study Author(s): Klinkert, A.

Testing Laboratory: Eurofins Agroscience Services Eco Chem GmbH

Eutinger Str. 24

75223 Niefern-Öschelbronn

Germany

Laboratory Report No.: 721406

Sponsor Study No.: BASF Reg. Doc. #: 2015/7001306

Study Completion Date: 24 November 2015

Data Access: Data submitter is data owner

Data Protection Claimed: Yes

II. MATERIALS AND METHODS

Test Guideline: Non-guideline

Deviations from Guideline: Calibration of boom sprayer conducted one day prior to application

rather than on day of application. During the calibration, first spray nozzle was outside of the 5% deviation of the mean (151.9 vs 160.0). Less than 0.5 g of pollen collected for several sampling dates and samples; however, residue analysis could be completed on the amount

of sample collected.

Storage temperature rose above -18oC on several occasions.

GLP Compliance: Study conducted compliant with the Principles of Good Laboratory

Practice (GLP) (Chemicals Act, Annex 1, Germany); the OECD Principles

of Good Laboratory Practice, which are accepted by regulatory

authorities throughout the European Community, the United States of America (EPA and FD) and Japan (MHW, MAFF and METI) on the basis of intergovernmental agreements. The analytical phase was compliant with EPA Good Laboratory Practice requirements (40 CFR 160).

A. MATERIALS

Test Material: BAS 440 00 I (Reg. no. 5 599 022)

Active ingredient: BAS 440 I (afidopyropen)

CAS Number 915972-17-7

Test Material Identity: Batch No. 17670104-1; dispersible concentrate; 9.7% (w/w) 100 g a.i./L.

Liquid/yellow.

Details on Preparation and Application of Test Materials:

Content of active ingredient: BAS 440 I: 9.7% w/w; (nominal: 100 g/L); batch no. 1767-104-1; density: 1.0235g/cm³. The test item solution was

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prepared shortly before the application. Homogeneity of the test item solution was obtained by thorough stirring or mixing. The application was carried out during bee flight at flowering of the crop (*Brassica* sp.). All substances were applied in 500 L/ha water using a calibrated

backpack boom sprayer.

Analytical Monitoring:
Details on Analytical Method:

Yes (see below)

Specimens of canola flowers, nectar from forager bees and pollen from forager bees were analyzed for residues of afidopyropen and its M4401007 degradate following the provisions of BASF Method D1412: 'Determination of Residues of BAS 440 I and its Metabolite M4401007 in Bee – Related Matrices by LCMS/MS' where residues of afidopyropen were extracted and partitioned using acetonitrile and salt solution, and then cleaned-up using a dispersive solid-phase extraction (dSPE) technique. The determination of afidopyropen was performed by high performance liquid chromatography (HPLC) positive ion electrospray ionization tandem mass spectrometry (MS/MS-ESI), monitoring ion transitions m/z 594→148 for BAS 440 I and m/z 1188→576 for the metabolite M4401007. The limit of quantification (LOQ) for afidopyropen and M4401007 in bee-related matrices was 0.01 mg/kg each. The method limit of detection (LOD) was estimated at 20% of the limit of quantitation, equivalent to 0.002 mg/kg for each of the analytes.

Mean recovery of M4401007 from flowers fortified at the LOQ was 66% (CV=10%); method-detector response was lintera over the 0.05-1.0 ng/mL range (r= 0.9915-0.9998); the limit of quantitation (LOQ), based on the lowest level of method validation (LLMV) was approximately 0.01 mg/kg for each analyte in the various matrices. Mean recoveries from flowers, pollen and nectar "shipping verification" sampels were 97-117% of nominal; field spikes were stored for up to 327 days prior to extraction.

Reference Material:

Reference Material Identity

Vehicle:

None N/A N/A

Test Organism (Species):

Apis mellifera L. (honeybee)

Animal Group:

Arthropoda/Insecta/Hymenoptera/Apidae

Details on Test Organisms:

Honeybees were only used as sampling device. No biological

observations were made. Honeybees, *Apis mellifera* L. (Hymenoptera, *Apidae*) of the race Italian bee, as typical for commercial use were used as sampling device for pollen and nectar. The colonies were bred by Eurofins Agroscience Services, Inc. For each tunnel, one honeybee hive was used. The hives contained ten frames with 4-5 frames containing pollen and honey and approximately 8000 bees in average. The hives

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were visibly healthy and queen-right. The hives were placed inside their respective tunnels in the morning of 05 Jun 2014 (BBCH: 10% 59, 10% 62, 40% 63, 20% 64, 10% 65, 10% 67) – one day before the first

sampling of pollen and forager bees.

B. STUDY DESIGN AND METHODS

Study Type: Semi-field (tunnel) study

Test Duration Type: Determination of residues of BAS 440 I and its metabolite M440I007 in

bee-relevant matrices up to 11 days after application of the test item

Limit Test: Yes
Total Exposure Duration: 11 days
Post-Exposure Observation Period:

Honey bees were only used as sampling device. No biological

observations were made

Test Location: This residue study was conducted in Cedar Grove, region Mebane,

North Carolina, USA

Test Environmental Conditions: Natural field conditions; during the application the temperature, wind

speed, cloud coverage and humidity data were recorded in the field. For the entire field phase, the following weather data were taken from the weather station at the Cedar Grove testing facility at a distance of approx. 200 m to the field site: daily min/max and mean temperature, daily min/max and mean humidity (GLP record). Daily precipitation was measured with a rain gauge placed in one of the tunnels (GLP). Air temperatures were at 19 °C - 30 °C before application, 16 °C - 28 °C during application of the test item, 14 °C - 33 °C during sampling phase.

50% – 95% (range; mean: 76%) during application

Photoperiod and Lighting: Natural (ambient) environmental conditions.

Nominal and Measured Concentrations:

Nominal: negative control (untreated water), 0.5 L BAS 440 00 I/ha (corresponding to 50 g BAS 440 I/ha); all treatments were applied in 500

L water/ha. Nominal treatment concentration was not verified

analytically.

Test Plots: Plots were planted with full flowering canola (*Brassica* sp.). The total

area covered per tunnel was approximately 137 m². The dimension of each tunnel was approximately 22.86 m in length, 6 m (Ta, C) in width and 2.8 m in height (at the center). A path of approx. 0.5 m was created by mulching the crop in the middle of the tunnel, because the spray width of the boom sprayer did not cover the middle part. The remaining crop area was approx. 125 m². The tunnel frames were covered with light plastic gauze. In each tunnel tent, one small bee colony was introduced one day before application and first sampling (in the morning of 05 Jun 2014, 1DBA). A container filled with water was placed

into each tunnel as water supply for the bees.

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Test Design:

Honey bees were used as sampling device for Canola pollen and nectar. Commercial honey bees were set up one day before application and first sampling. Samples of honeybee-collected pollen, nectar prepared from forager bees and canola flowers were taken for residue analysis 4 times (DAAO, 3, 7 and 10) until end of flowering, to be subsequently analyzed for residues of BAS 440 I (afidopyropen) and its metabolite M440I007.

Sampling of Canola inflorescences: The terminal inflorescences with open flowers were cut at the end of the stipe and hand bagged. The samples of flowers were collected from at least 12 different locations across the plot, respectively. Flowers were sampled 4 times during flowering period (approx. BBCH 62-67 to BBCH 69). Samples were divided into sub-sample A (with at least 5 g) for residue analysis and R as retained sample. The sampling always started in the control group or was conducted by different personnel.

Sampling of pollen from pollen traps: The honeybee hives were equipped with pollen traps as used for commercial pollen collection. At the sampling day the grid of the pollen trap was inserted during time of foraging activity of the bees and is kept in place for approximately 4 - 8 hours from when inserted. After this period, the grid was removed from the pollen trap and the collected pollen was sampled. Samples were divided into sub-sample A (with at least 0.5 g) for residue analysis and R as retained sample. The sampling always started in the control group or was conducted by different personnel.

Sampling of pollen from inflorescences (additional sample): Additional pollen was manually collected from flowers on ODAA in the test item treatment group. Therefore, flowers were rubbed against a sieve and pollen which had fallen through the sieve was collected.

Sampling of forager bees for preparation of nectar: Shortly before sampling, the hive entrances were sealed and the forager bees were subsequently collected as they returned to the hive by collecting them into a box containing dry ice, either by using modified hoovers ("bee vac"). Directly after sampling, the sample was split in two sub-samples (A for residue analysis (≥ 300 bees) and retained sample (R-sample) (≥150 bees, or all remaining)). If less forager bees were available, the R sample contains less forager bees. The sampling always started in the control group or was conducted by different personnel.

<u>Nectar from forager bees for residue analysis</u>: All forager bee samples (A and R samples) were prepared from the honey stomach and the nectar of the respective subsamples was pooled. The weight of the

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pooled samples was recorded. The nectar samples were unfrozen for <2 hours during preparation and were transferred back to the freezer immediately afterwards.

After collection of the specimens, flower samples were deep frozen 5 min after end of sampling (ambient transport to freezer, 5 min transport) and subsequently stored deep frozen until residue analysis.

Pollen from pollen traps and inflorescences: Samples were transported on dry ice and subsequently deep frozen until residue analysis. Sampling of forager bees for preparation of nectar: Samples were transported on dry ice and subsequently deep frozen until residue analysis.

III. APPLICANT'S REPORTED RESULTS AND DISCUSSION

Exposure Duration: 11 days

Endpoint(s): Residues of afidopyropen and its metabolite M440I007 in inflorescences

of Canola (*Brassica* sp.) as well as in pollen collected with pollen traps and nectar prepared from honey stomachs of forager honeybees after exposure to flowering *Brassica* sp., treated once with BAS 440 00 I during flowering in a semi-field residue study in North Carolina. The treated samples in this study were stored frozen for 275 - 327 days, from collection to extraction for analysis (approximately 9 - 11 months). According to the study authors, the available storage stability data from the shipping verification samples (field spikes performed in conjunction

with the subject study) supported the storage intervals.

Basis for Concentration: Nominal
Effect Concentration Type: Test material

Basis for Effect: Measured residues in flowers and bee-collected pollen/nectar from

treated canola (Brassica spp).

Applicant-Provided Results:

The study report indicated recoveries of afidopyropen in flowers spiked with 0.01, 1 and 5 mg/kg averaged 98%, while recoveries of M4401007 averaged 73%. Recoveries of afidopyropen from nectar spiked with 0.01 or 1.0 mg/kg averaged 96% while recoveries of M4401007 averaged 82%. Recoveries of afidopyropen from pollen spiked with 0.01 or 1.0 mg/kg averaged 97% while M4401007 averaged 84%. Recoveries of field spikes of 1.0 mg/kg fanged between 97 – 117% across all of the matrices.

According to the study authors, mean residues of afidopyropen in flowers were 4.43 mg/kg (range: 3.73 - 4.97 mg/kg) at 0 days after application (0 DAA), declined to 0.11 mg/kg (range: 0.10 - 0.13 mg/kg) at 3 DAA, and 0.013 mg/kg (range: 0.011 - 0.015 mg/kg) at 7 DAA, and were non-quantifiable (<0.01 mg/kg) at 10 DAA (**Table 1**). Mean M440I007 residues in flowers were 0.36 mg/kg (range: 0.33 - 0.40 mg/kg) at 0 days after treatment (0 DAA), declined to 0.020 mg/kg (range: 0.014 - 0.023 mg/kg) at 3 DAA, and were non-quantifiable (<0.01 mg/kg) in all samples collected 7 and 10 days after treatment.

The study authors reported that in pollen, mean residues of afidopyropen were 0.26 mg/kg (range: 0.17 - 0.40 mg/kg) at 0 DAA, were 0.023 mg/kg (range, 0.018 - 0.031 mg/kg) and 0.027 mg/kg (range: 0.020 -

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0.037 mg/kg) at 3 and 7 DAA, respectively, and declined to <0.010 mg/kg (range: <0.010 -0.011 mg/kg) at 10 DAA (**Table 1**). Mean M440I007 residues in pollen were 0.061 mg/kg (range: 0.037 - 0.10 mg/kg) at 0 DAA and were non-quantifiable (each replicate <0.01 mg/kg, except one at 0.018 mg/kg at 3 DAA) for the remaining sampling intervals.

According to the study authors, residues of afidopyropen in bee-collected nectar were 0.052 mg/kg (range: 0.010 - 0.13 mg/kg) at 0 DAA, and were below the limit of quantitation (<0.01 mg/kg) in all samples collected 3, 7, and 10 days after treatment (**Table 1**). Mean M440l007 residues in nectar were below the limit of quantitation (each replicate <0.01 mg/kg except one at 0.013 mg/kg at 0 DAA) in all samples collected 0, 3, 7, and 10 days after treatment.

Table 1. Registrant summary of measured afidopyropen parent and M4401007 degradate residues (range and mean) in flowers, and honey bee (*Apis mellifera*)-collected pollen and nectar from canola (*Brassica* spp) at 0, 3, 7 and 10 days after application of afidopyropen typical end product BAS 440 00 I (9.7% active ingredient) during bloom.

		Residue levels [mg/kg] 1)					
Analyte (Matrix)	Days after treatment	ment n (afidopyrop			M440I007		
	(DAA)		Range	Mean	Range	Mean	
	0	3	3.73 - 4.97	4.43	0.33 - 0.40	0.36	
- Classiana	3	3	0.10 - 0.13	0.11	0.01 - 0.02	0.02	
Flowers	7	3	0.01 - 0.02	0.01	< 0.01	< 0.01	
	10	3	< 0.01	< 0.01	< 0.01	< 0.01	
	0	3	0.17 - 0.40	0.26	0.04 - 0.10	0.06	
Deller	3	3	0.02 - 0.03	0.02	< 0.01 - 0.02	< 0.01	
Pollen	7	3	0.02 - 0.04	0.03	< 0.01	< 0.01	
	10	3	<0.01 - 0.01	< 0.01	< 0.01	< 0.01	
	0	3	0.01 - 0.13	0.05	< 0.01 - 0.01	< 0.01	
Nastau	3	3	< 0.01	< 0.01	< 0.01	< 0.01	
Nectar	7	3	< 0.01	< 0.01	< 0.01	< 0.01	
	10	3	< 0.01	< 0.01	< 0.01	< 0.01	

 $^{^1}$ Values < 0.01 mg/kg were assigned ½ the limit of quantification (LOQ) of 0.005 mg/kg for calculation of the mean.

Applicant-Reported Statistics and Error Estimates

Statistical treatment of the data included simple descripting statistics (e.g., mean; range) for the residue and method recovery data, and calculation of the calibration curve and coefficient of variation (r) by linear regression of the instrument responses for the reference standards.

IV. OVERALL REMARKS, ATTACHMENTS

Multiple deviations from the study protocol were noted by the author; however, in general, the study authors did not believe these deviations impacted the scientific integrity of the study.

V. PRIMARY REVIEWER'S ANALYSIS AND CONCLUSIONS

Average residues of parent afidopyropen and its M440I007 degradate are consistent with those calculated by the study authors and reported in **Table 1**.

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Reviewer's Statistical Verification:

Descriptive statistics on residue levels at various study sampling intervals were generated using the Proc Univariate procedure of SAS® (SAS Institute, Cary, NC).

Reviewer's Comments:

According to the study report, an investigation of the storage stability of residues (afidopyropen and M4410007) frozen in a variety of plant matrices is currently in progress; however, the report notes that "available data indicate that residues of afidopyropen are stable when stored frozen in representative crop matrices for at least 12 months." Nominal concentration in application solution was not verified analytically.

Table 2 summarizes the weight of pollen collected for each of the study replicates and indicates that at 0 DAA, the weight of pollen collected in sample A controls ($C_a - C_b$) averaged 1.188 g; whereas, the weight of pollen collected in afidopyropen treatments ($T_a - T_c$) for sample A averaged 0.271 g and were roughly 77% lower than controls even though pollen trap samples had been supplemented with pollen collected directly from flowers. At 3 DAA, control and treated sample weights averaged 3.064 and 0.687 g, respectively, in sample A; weights were again roughly 78% lower than controls. By 7 DAA, mean pollen sample weights in control and treated averaged, 0.548 and 0.757 g, respectively, in sample A; by this time, pollen weights had increased by roughly 38% relative to controls. At 10 DAA, mean pollen weights in control and treated averaged 0.535 and 0.729 g, respectively; by this time, pollen weights from bees foraging in treated areas increased by roughly 36% relative to controls.

Table 2. Summary of pollen sample weights for each replicate of control (C) and afidopyropen treated (T) at different sampling interval (days after treatment; DAA).

Doubleate	Sample	BBCII	Sample V	Veight (g)
Replicate	Interval	ВВСН	Subsample A	Subsample B
Ca			1.193	5.491
C _b		10% 59,	1.182	0.505
Ta	0 DAA	80% 62 – 65	0.267ª	N/A
T _b		10% 67	0.317ª	N/A
T _c			0.229ª	N/A
Ca		100/.62	2.187	1.704
C _b]	10% 62	3.940	4.044
Ta	3 DAA	60% 63 – 65 20% 67	0.530	0.586
T _b]		0.562	0.423
T _c	†	10% 69	0.968	0.539
Ca			0.589	0.219
Сь	1	30% 65	0.506	0.069
Ta	7 DAA	40% 67	0.566	N/A
T _b]	30% 69	1.246	0.506
T _c]		0.459	N/A
Ca			0.497	0.046
Сь	10 DAA	20% 69	0.573	0.857
T _a] IU DAA	80% 69	0.516	N/A
T _b]		1.114	0.508

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T _c		0.557	N/A	

DAA = days after application

N/A = not applicable

Following a foliar spray application of the afidopyropen formulated product BAS 440 00 I (9.7% a.i.) at a nominal rate of 50 g BAS 440 I/ha (0.272 lbs/A) to canola at bloom mean afidopyropen residues in flowers were 4.43 mg/kg (range: 3.73 - 4.97 mg/kg) at 0 days after application (0 DAA), 0.11 mg/kg (range: 0.10 - 0.13 mg/kg) at 3 DAA, and 0.013 mg/kg (range: 0.011 - 0.015 mg/kg) at 7 DAA, and were less than the limit of quantification (LOQ<0.01 mg/kg) at 10 DAA. Mean M440I007 residues were 0.36 mg/kg (range: 0.33 - 0.40 mg/kg) at 0 DAA, 0.020 mg/kg (range: 0.014 - 0.023 mg/kg) at 3 DAA, and were below the LOQ (<0.01 mg/kg) in all samples collected 7 and 10 DAA.

In pollen, mean afidopyropen residues were 0.26 mg/kg (range: 0.17 - 0.40 mg/kg) at 0 DAA, were 0.023 mg/kg (range, 0.018 - 0.031 mg/kg) and 0.027 mg/kg (range: 0.020 - 0.037 mg/kg) at 3 and 7 DAA, respectively, and declined to <0.010 mg/kg (range: <0.010 - 0.011 mg/kg) at 10 DAA. Mean M4401007 residues were 0.061 mg/kg (range: 0.037 - 0.10) at 0 DAA and were below the LOQ (each replicate <0.01 mg/kg, except one at 0.018 mg/kg at 3 DAA) for the remaining sampling intervals.

In nectar, mean afidopyropen residues were 0.052 mg/kg (range: 0.010 - 0.13 mg/kg) at 0 DAA, and were below the LOQ (<0.01 mg/kg) in all samples collected 3, 7, and 10 DAA. Mean M440I007 residues were below the LOQ (each replicate <0.01 mg/kg except one at 0.013 mg/kg at 0 DAA) in all samples collected 0, 3, 7, and 10 DAA.

Reviewer's Conclusions:

Following a foliar spray application of the afidopyropen formulated product BAS 440 00 I (9.7% a.i.) at a nominal rate of 50 g BAS 440 I/ha (0.272 lbs/A) to canola at bloom mean (\pm standard deviation) afidopyropen residues in flowers were 4.43 mg/kg (\pm 0.63) at 0 days after application (0 DAA), 0.11 mg/kg (\pm 0.015) at 3 DAA, and 0.013 mg/kg (\pm 0.002) at 7 DAA, and were less than the limit of quantification (LOQ<0.01 mg/kg) at 10 DAA. Mean M440I007 residues were 0.36 mg/kg (\pm 0.035) at 0 DAA, 0.020 mg/kg (\pm 0.005) at 3 DAA, and were below the LOQ (<0.01 mg/kg) in all samples collected 7 and 10 DAA.

In pollen, mean afidopyropen residues were 0.26 mg/kg (\pm 0.12) at 0 DAA, were 0.023 mg/kg (\pm 0.027) and 0.027 mg/kg (\pm 0.009) at 3 and 7 DAA, respectively, and declined to <0.010 mg/kg (range: <0.010 - 0.011 mg/kg) at 10 DAA. Mean M440I007 residues were 0.061 mg/kg (range: 0.037 - 0.10) at 0 DAA and were below the LOQ (each replicate <0.01 mg/kg, except one at 0.018 mg/kg at 3 DAA) for the remaining sampling intervals.

In nectar, mean afidopyropen residues were 0.052 mg/kg (±0.068) at 0 DAA, and were below the LOQ (<0.01 mg/kg) in all samples collected 3, 7, and 10 DAA. Mean M440I007 residues were below the LOQ (each replicate <0.01 mg/kg except one at 0.013 mg/kg at 0 DAA) in all samples collected 0, 3, 7, and 10 DAA.

Although the study was not intended to provide biological information on the bees used to collect samples, weights of pollen samples provide anecdotal information that at 0 and 3 DAA, samples sizes from treated plots were 77 – 78% lower than controls; however, by 7 and 10 DAA, pollen sample

^a additional pollen collected from flowers in T_a, T_b and T_c

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weights from treated plots had increased by 38 and 36%, respectively, relative to controls. These data suggest that bee pollen foraging activity may have been affected in afidopyropen-treated plots from 0 through 3 DAA; however, after this period, bee pollen foraging activity may have increased in the treated group relative to controls. Based on sample sizes of nectar, nectar foraging activity did not appear to be affected.

Results Synopsis:

Mean residues at 0 DAA after foliar application of BAS 440 00 I at nominal rate of 50 g a.i./ha (0.272 lbs a.i./A):

Flowers: 4.43 ± 0.63 mg/kg (Parent); 0.36 ± 0.035 mg/kg (M4401007) Nectar: 0.052 ± 0.068 mg/kg (Parent); <0.01 mg/kg (M4401007) Pollen: 0.26 ± 0.12 mg/kg (Parent); 0.061 ± 0.034 mg/kg (M4401007)

EPA Classification: Acceptable

PMRA Classification: Reliable with restrictions

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APPENDIX I. Output of Statistics Verified by the Reviewer

DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DEGRADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure Variable: PARENT SAMPLE=Flower DAA=0

Moments

N	3	Sum Weights	3
Mean	4.42666667	Sum Observations	13.28
Std Deviation	0.63406099	Variance	0.40203333
Skewness	-1.0245835	Kurtosis	
Uncorrected SS	59.5902	Corrected SS	0.80406667
Coeff Variation	14.3236668	Std Error Mean	0.36607528

Basic Statistical Measures

Location		Variability	
Mean	4.426667	Std Deviation	0.63406
Median	4.580000	Variance	0.40203
Mode	•	Range	1.24000
		Interquartile Range	1.24000

Tests for Location: Mu0=0

Test	Statistic		p Value	
Student's t	t	12.09223	Pr > t	0.0068
Sign	М	1.5	Pr >= M	0.2500
Signed Rank	S	3	Pr >= S	0.2500

Quantiles (Definition 5)

Level	Quantile
100% Max	4.97
99%	4.97
95%	4.97
90%	4.97
75% Q3	4.97
50% Median	4.58
25% Q1	3.73
10%	3.73
5%	3.73
1%	3.73

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Quantiles (Definition 5)

Level Quantile 0% Min 3.73

Extreme Observations

 Lowest
 Highest

 Value
 Obs
 Value
 Obs

 3.73
 2
 3.73
 2

 4.58
 3
 4.58
 3

 4.97
 1
 4.97
 1

DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DE RADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure Variable: DEGRADATE SAMPLE=Flower DAA=0

Moments

Ν 3 **Sum Weights** 3 Mean 0.36333333 Sum Observations 1.09 Std Deviation 0.03511885 Variance 0.00123333 Skewness 0.42327316 Kurtosis Uncorrected SS 0.3985 **Corrected SS** 0.00246667 Coeff Variation 9.66573739 Std Error Mean 0.02027588

Basic Statistical Measures

 Mean
 0.363333
 Std Deviation
 0.03512

 Median
 0.360000
 Variance
 0.00123

 Mode
 .
 Range
 0.07000

 Interquartile Range
 0.07000

Tests for Location: Mu0=0

 Test
 Statistic
 p Value

 Student's t
 t 17.91949
 Pr > |t| 0.0031

 Sign
 M 1.5
 Pr >= |M| 0.2500

 Signed Rank
 S 3
 Pr >= |S| 0.2500

Quantiles (Definition 5)

Level Quantile **100% Max** 0.40

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Quantiles (Definition 5)				
Quantile				
0.40				
0.40				
0.40				
0.40				
0.36				
0.33				
0.33				
0.33				
0.33				
0.33				

Extreme Observations

Lowes	t	Highest		
Value	Obs	Value	Obs	
0.33	2	0.33	2	
0.36	3	0.36	3	
0.40	1	0.40	1	

DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DE RADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure

Variable: PARENT SAMPLE=Flower DAA=3

Moments

N	3	Sum Weights	3
Mean	0.11333333	Sum Observations	0.34
Std Deviation	0.01527525	Variance	0.00023333
Skewness	0.93521953	Kurtosis	
Uncorrected SS	0.039	Corrected SS	0.00046667
Coeff Variation	13.4781638	Std Error Mean	0.00881917

Basic Statistical Measures

Location			Variability		
	Mean	0.113333	Std Deviation	0.01528	
	Median	0.110000	Variance	0.0002333	
	Mode	•	Range	0.03000	
			Interquartile Range	0.03000	

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Tests for Location: Mu0=0

 Test
 Statistic
 p Value

 Student's t
 t
 12.85079
 Pr > |t|
 0.0060

 Sign
 M
 1.5
 Pr >= |M|
 0.2500

 Signed Rank
 S
 3
 Pr >= |S|
 0.2500

Quantiles (Definition 5)

Level	Quanti
100% Max	0.13
99%	0.13
95%	0.13
90%	0.13
75% Q3	0.13
50% Median	0.11
25% Q1	0.10
10%	0.10
5%	0.10
1%	0.10
0% Min	0.10

Extreme Observations

Lowes	t	Highest		
Value	Obs	Value	Obs	
0.10	4	0.10	4	
0.11	6	0.11	6	
0.13	5	0.13	5	

DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DEGRADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure Variable: DEGRADATE SAMPLE=Flower DAA=3

Moments

N	3	Sum Weights	3
Mean	0.01966667	Sum Observations	0.059
Std Deviation	0.00493288	Variance	0.00002433
Skewness	-1.6523167	Kurtosis	
Uncorrected SS	0.001209	Corrected SS	0 00004867

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Moments

Coeff Variation 25.0824552 Std Error Mean 0.002848

Basic Statistical Measures

Location		Variability		
Mean	0.019667	Std Deviation	0.00493	
Median	0.022000	Variance	0.0000243	
Mode	•	Range	0.00900	
		Interquartile Range	0.00900	

Tests for Location: Mu0=0

Test	Statistic		p Value	
Student's t	t	6.905428	Pr > t	0.0203
Sign	М	1.5	Pr >= M	0.2500
Signed Rank	S	3	Pr >= S	0.2500

Quantiles (Definition 5)

Level	Quantile
100% Max	0.023
99%	0.023
95%	0.023
90%	0.023
75% Q3	0.023
50% Median	0.022
25% Q1	0.014
10%	0.014
5%	0.014
1%	0.014
0% Min	0.014

Extreme Observations

Lowest		Highest		
Value	Obs	Value	Obs	
0.014	4	0.014	4	
0.022	6	0.022	6	
0.023	5	0.023	5	

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The UNIVARIATE Procedure

Variable: PARENT SAMPLE=Flower DAA=7

Moments

N	3	Sum Weights	3
Mean	0.01266667	Sum Observations	0.038
Std Deviation	0.00208167	Variance	4.33333E-6
Skewness	1.29334278	Kurtosis	
Uncorrected SS	0.00049	Corrected SS	8.66667E-6
Coeff Variation	16.4342053	Std Error Mean	0.00120185

Basic Statistical Measures

Location		Variability		
Mean	0.012667	Std Deviation	0.00208	
Median	0.012000	Variance	4.33333E-6	
Mode	•	Range	0.00400	
		Interquartile Range	0.00400	

Tests for Location: Mu0=0

Test	Sta	itistic	p Value	
Student's t	t	10.5393	Pr > t	0.0089
Sign	М	1.5	Pr >= M	0.2500
Signed Rank	S	3	Pr >= S	0.2500

Quantiles (Definition 5)

Level	Quantile
100% Max	0.015
99%	0.015
95%	0.015
90%	0.015
75% Q3	0.015
50% Median	0.012
25% Q1	0.011
10%	0.011
5%	0.011
1%	0.011
0% Min	0.011

Extreme Observations

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 Lowest
 Highest

 Value
 Obs
 Value
 Obs

 0.011
 7
 0.011
 7

 0.012
 9
 0.012
 9

 0.015
 8
 0.015
 8

DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DE RADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure Variable: DEGRADATE SAMPLE=Flower DAA=7

Missing Values

Missing Count Percent Of

Value

All Obs Missing Obs

3 100.00 100.00

DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DEGRADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure

Variable: PARENT

SAMPLE=Flower DAA=10

Missing Values

Missing Count Percent Of

Value

All Obs Missing Obs

3 100.00 100.00

DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DEGRADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure Variable: DEGRADATE SAMPLE=Flower DAA=10

Missing Values

Missing Count Percent Of

Value

All Obs Missing Obs

3 100.00 100.00

DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DEGRADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure Variable: PARENT SAMPLE=Nectar DAA=0

Moments

N 3 Sum Weights 3

Mean 0.05166667 **Sum Observations** 0.155

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M	n	m	e	n	ts

Std Deviation 0.06788471 **Variance** 0.00460833

Skewness 1.72148595 Kurtosis

 Uncorrected SS
 0.017225
 Corrected SS
 0.00921667

 Coeff Variation
 131.389754
 Std Error Mean
 0.03919325

Basic Statistical Measures

Location		variability	
Mean	0.051667	Std Deviation	0.06788
Median	0.015000	Variance	0.00461
Mode	•	Range	0.12000
		Interquartile Range	0.12000

Tests for Location: Mu0=0

Test	Sta	tistic	p Value	
Student's t	t	1.318254	Pr > t	0.3181
Sign	М	1.5	Pr >= M	0.2500
Signed Rank	S	3	Pr >= S	0.2500

Quantiles (Definition 5)

Level	Quantile
100% Max	0.130
99%	0.130
95%	0.130
90%	0.130
75% Q3	0.130
50% Median	0.015
25% Q1	0.010
10%	0.010
5%	0.010
1%	0.010
0% Min	0.010

Extreme Observations

Lowest			Highest		
Val	ue	Obs	Value	Obs	
0.0	10	14	0.010	14	
0.03	15	13	0.015	13	

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Extreme Observations

Value Obs Value Obs
0.130 15 0.130 15

DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DE RADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure Variable: DEGRADATE SAMPLE=Nectar DAA=0

Moments

N1Sum Weights1Mean0.013Sum Observations0.013Std Deviation.Variance.Skewness.Kurtosis.Uncorrected SS0.000169Corrected SS0Coeff Variation.Std Error Mean.

Basic Statistical Measures

LocationVariabilityMean0.013000Std Deviation.Median0.013000Variance.Mode0.013000Range0Interquartile Range0

Tests for Location: Mu0=0

Test Statistic p Value
Student's t t . Pr > |t| .

Sign M 0.5 $Pr \ge |M|$ 1.0000 Signed Rank S 0.5 $Pr \ge |S|$ 1.0000

Quantiles (Definition 5)

Level	Quantile
100% Max	0.013
99%	0.013
95%	0.013
90%	0.013
75% Q3	0.013
50% Median	0.013

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Quantiles (Definition 5)

Quantile Level 25% Q1 0.013 10% 0.013 5% 0.013 0.013 1% 0% Min 0.013

Extreme Observations

Lowest Highest Value Obs Value Obs 0.013 15 0.013 15

Missing Values

Missing Count Percent Of

Value

All Obs Missing Obs

66.67 100.00

DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DEGRADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure Variable: PARENT

SAMPLE=Nectar DAA=3

Missing Values

Missing Count Percent Of

Value

All Obs Missing Obs

100.00 100.00

DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DEGRADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure Variable: DEGRADATE SAMPLE=Nectar DAA=3

Missing Values

Missing Count Percent Of

Value

All Obs Missing Obs

3 100.00 100.00

DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DEGRADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure

Variable: PARENT

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SAMPLE=Nectar DAA=7

Missing Values

Missing Count Percent Of

Value

All Obs Missing Obs

3 100.00 100.00

DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DEGRADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure Variable: DEGRADATE SAMPLE=Nectar DAA=7

Missing Values

Missing Count Percent Of

Value

All Obs Missing Obs

3 100.00 100.00

DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DEGRADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure

Variable: PARENT SAMPLE=Nectar DAA=10

Missing Values

Missing Count Percent Of

Value

All Obs Missing Obs

3 100.00 100.00

DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DEGRADATE IN VARIOUS MATRICES.

The UNIVARIATE Procedure Variable: DEGRADATE SAMPLE=Nectar DAA=10

Missing Values

Missing Count Percent Of

Value

All Obs Missing Obs

. 3 100.00 100.00

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DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DEGRADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure

Variable: PARENT SAMPLE=Pollen DAA=0

Moments

N	3	Sum Weights	3
Mean	0.26333333	Sum Observations	0.79
Std Deviation	0.12096832	Variance	0.01463333
Skewness	1.40513837	Kurtosis	
Uncorrected SS	0.2373	Corrected SS	0.02926667
Coeff Variation	45 937335	Std Frror Mean	0 06984109

Basic Statistical Measures

Location		Variability	
Mean	0.263333	Std Deviation	0.12097
Median	0.220000	Variance	0.01463
Mode		Range	0.23000
		Interquartile Range	0.23000

Tests for Location: Mu0=0

Test	Sta	itistic	p Value	
Student's t	t	3.770464	Pr > t	0.0637
Sign	М	1.5	Pr >= M	0.2500
Signed Rank	S	3	Pr >= S	0.2500

Quantiles (Definition 5)

Level	Quantil
100% Max	0.40
99%	0.40
95%	0.40
90%	0.40
75% Q3	0.40
50% Median	0.22
25% Q1	0.17
10%	0.17
5%	0.17
1%	0.17

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Quantiles (Definition 5)

Level Quantile 0% Min 0.17

Extreme Observations

 Lowest
 Highest

 Value
 Obs
 Value
 Obs

 0.17
 26
 0.17
 26

 0.22
 25
 0.22
 25

 0.40
 27
 0.40
 27

DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DE RADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure Variable: DEGRADATE SAMPLE=Pollen DAA=0

Moments

N	3	Sum Weights	3
Mean	0.06066667	Sum Observations	0.182
Std Deviation	0.03429772	Variance	0.00117633
Skewness	1.62663966	Kurtosis	
Uncorrected SS	0.013394	Corrected SS	0.00235267
Coeff Variation	56.534697	Std Error Mean	0.0198018

Basic Statistical Measures

Location		Variability	
Mean	0.060667	Std Deviation	0.03430
Median	0.045000	Variance	0.00118
Mode	•	Range	0.06300
		Interquartile Range	0.06300

Tests for Location: Mu0=0

Test	Sta	ntistic	p Value	
Student's t	t	3.063695	Pr > t	0.0921
Sign	М	1.5	Pr >= M	0.2500
Signed Rank	s	3	Pr >= S	0.2500

Quantiles (Definition 5)

Level	Quantile
100% Max	0.100

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Quantiles (Definition 5)		
Level	Quantile	
99%	0.100	
95%	0.100	
90%	0.100	
75% Q3	0.100	
50% Median	0.045	
25% Q1	0.037	
10%	0.037	
5%	0.037	
1%	0.037	
0% Min	0.037	
Extreme Obs	ervations	
Lowest	Highest	
Value Obs	Value Obs	

DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DE RADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure

Variable: PARENT SAMPLE=Pollen DAA=3

Moments

N	3	Sum Weights	3
Mean	0.023	Sum Observations	0.069
Std Deviation	0.007	Variance	0.000049
Skewness	1.57434402	Kurtosis	
Uncorrected SS	0.001685	Corrected SS	0.000098
Coeff Variation	30.4347826	Std Error Mean	0.00404145

0.037 26 0.037 26 0.045 25 0.045 25 0.100 27 0.100 27

Basic Statistical Measures

Location			Variability		
	Mean	0.023000	Std Deviation	0.00700	
	Median	0.020000	Variance	0.0000490	
	Mode		Range	0.01300	
			Interquartile Range	0.01300	

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Tests for Location: Mu0=0

 Test
 Statistic
 p Value

 Student's t
 t
 5.691024
 Pr > |t|
 0.0295

 Sign
 M
 1.5
 Pr >= |M|
 0.2500

 Signed Rank
 S
 3
 Pr >= |S|
 0.2500

Quantiles (Definition 5)

Level	Quanti
100% Max	0.031
99%	0.031
95%	0.031
90%	0.031
75% Q3	0.031
50% Median	0.020
25% Q1	0.018
10%	0.018
5%	0.018
1%	0.018
0% Min	0.018

Extreme Observations

Lowes	t	Highest		
Value	Obs	Value	Obs	
0.018	28	0.018	28	
0.020	30	0.020	30	
0.031	29	0.031	29	

DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DEGRADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure Variable: DEGRADATE SAMPLE=Pollen DAA=3

Moments

N	1	Sum Weights	1
Mean	0.018	Sum Observations	0.018
Std Deviation		Variance	
Skewness		Kurtosis	
Uncorrected SS	0.000324	Corrected SS	0

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Moments

Coeff Variation . Std Error Mean .

Basic Statistical Measures

LocationVariabilityMean0.018000Std Deviation.Median0.018000Variance.Mode0.018000Range0Interquartile Range0

Tests for Location: Mu0=0

 Test
 Statistic p Value

 Student's t
 t
 ...
 Pr > |t|
 ...

 Sign
 M
 0.5
 Pr >= |M|
 1.0000

 Signed Rank
 S
 0.5
 Pr >= |S|
 1.0000

Quantiles (Definition 5)

Level	Quantile
100% Max	0.018
99%	0.018
95%	0.018
90%	0.018
75% Q3	0.018
50% Median	0.018
25% Q1	0.018
10%	0.018
5%	0.018
1%	0.018
0% Min	0.018

Extreme Observations

 Value
 Obs
 Value
 Obs

 0.018
 29
 0.018
 29

Missing Values

Missing Count Percent Of
Value All Obs Missing Obs

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Missing Values

Missing Count Percent Of

Value

All Obs Missing Obs

. 2 66.67 100.00

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DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DEGRADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure

Variable: PARENT SAMPLE=Pollen DAA=7

Moments

N	3	Sum Weights	3
Mean	0.02666667	Sum Observations	0.08
Std Deviation	0.00907377	Variance	0.00008233
Skewness	1.52149166	Kurtosis	
Uncorrected SS	0.002298	Corrected SS	0.00016467
Coeff Variation	34.026644	Std Error Mean	0.00523874

Basic Statistical Measures

Location			Variability		
	Mean	0.026667	Std Deviation	0.00907	
	Median	0.023000	Variance	0.0000823	
	Mode	•	Range	0.01700	
			Interquartile Range	0.01700	

Tests for Location: Mu0=0

Test	Sta	itistic	p Value	
Student's t	t	5.090278	Pr > t	0.0365
Sign	М	1.5	Pr >= M	0.2500
Signed Rank	S	3	Pr >= S	0.2500

Quantiles (Definition 5)

Level	Quanti
100% Max	0.037
99%	0.037
95%	0.037
90%	0.037
75% Q3	0.037
50% Median	0.023
25% Q1	0.020
10%	0.020
5%	0.020
1%	0.020

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Quantiles (Definition 5)

Level Quantile

0% Min 0.020

Extreme Observations

 Lowest
 Highest

 Value
 Obs

 0.020
 31
 0.020
 31

 0.023
 32
 0.023
 32

 0.037
 33
 0.037
 33

DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DE RADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure Variable: DEGRADATE SAMPLE=Pollen DAA=7

Missing Values

Missing Count Percent Of

Value

All Obs Missing Obs

. 3 100.00 100.00

DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENT AND DEGRADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure

Variable: PARENT

SAMPLE=Pollen DAA=10

Moments

N	2	Sum Weights	2
Mean	0.0105	Sum Observations	0.021
Std Deviation	0.00070711	Variance	5E-7
Skewness		Kurtosis	
Uncorrected SS	0.000221	Corrected SS	5E-7
Coeff Variation	6.7343503	Std Error Mean	0.0005

Basic Statistical Measures

Location Variability

 Mean
 0.010500
 Std Deviation
 0.0007071

 Median
 0.010500
 Variance
 5E-7

 Mode
 .
 Range
 0.00100

 Interquartile Range
 0.00100

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Tests for Location: Mu0=0

Test Statistic p Value

 Student's t
 t
 21
 Pr > |t| 0.0303

 Sign
 M
 1
 Pr >= |M| 0.5000

 Signed Rank
 S
 1.5
 Pr >= |S| 0.5000

Quantiles (Definition 5)

Level	Quantile
100% Max	0.0110
99%	0.0110
95%	0.0110
90%	0.0110
75% Q3	0.0110
50% Median	0.0105
25% Q1	0.0100
10%	0.0100
5%	0.0100
1%	0.0100
0% Min	0.0100

Extreme Observations

 Value
 Obs
 Value
 Obs

 0.010
 35
 0.010
 35

 0.011
 34
 0.011
 34

Missing Values

Missing Count Percent Of

Value

All Obs Missing Obs

1 33.33 100.00

DESCRIPTIVE STATISTICS FOR RESIDUES OF PARENTAL D DEGRADATE IN VARIOUS MATRICES

The UNIVARIATE Procedure Variable: DEGRADATE SAMPLE=Pollen DAA=10

Missing Values

Missing Count Percent Of

Value All Obs Missing Obs

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Missing Values

Missing Count Percent Of

Value

All Obs Missing Obs

. 3 100.00 100.00